## Characterisation of proteins

Stephan Uebel, Microchemistry Core Facility, MPI of Biochemistry

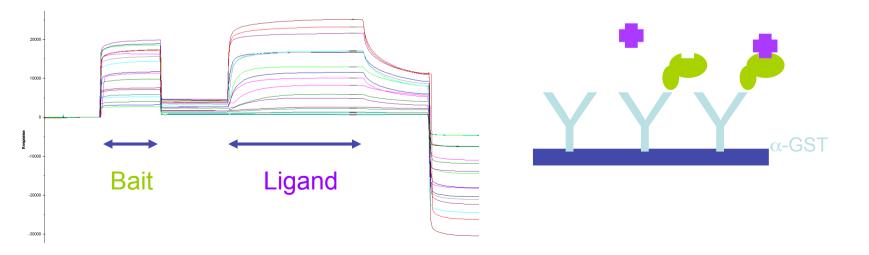
# Proteincharacterization – Why?

### Proteincharacterization is done for peace of mind

The key to a good Itc experiment is buffer matching andprotein quality Ernesto Frère @ DIPIA2010

> Sample quality is key to interaction proteomics success Ann-Claude Gingras, Mount Sinai Hospital

What are the requirements for performing high quality quantitative SPR experiments? 1. Learn how to recognize good quality data and 2. Prepare high quality samples, i.e. monodisperse and free of aggregates) Eileen M. Lafer, UTHSCSA Center for Macromolecular Interactions



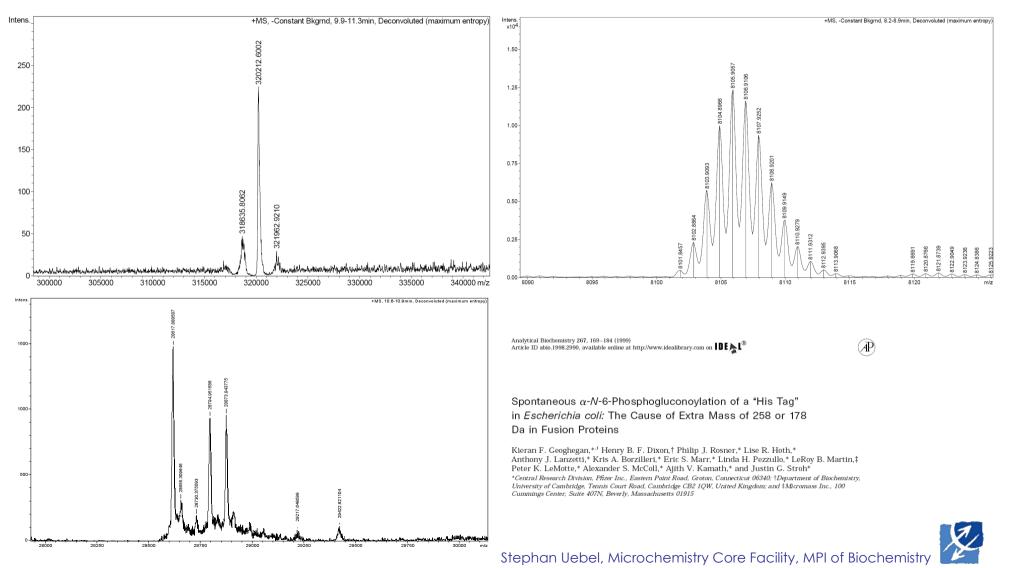


- Identitiv i.e. primary structure: amino acid sequence and desired posttranslational modifications
  - Edman sequencing combined with protein total mass (ESI-TOF, ESI-FTMS, [MALDI(IR)-TOF], deglycosylation of secreted proteins from insect cells and S.cerevisiae)
  - [peptide mapping by MS of tryptic digests]



## Resolution in Total Protein MS

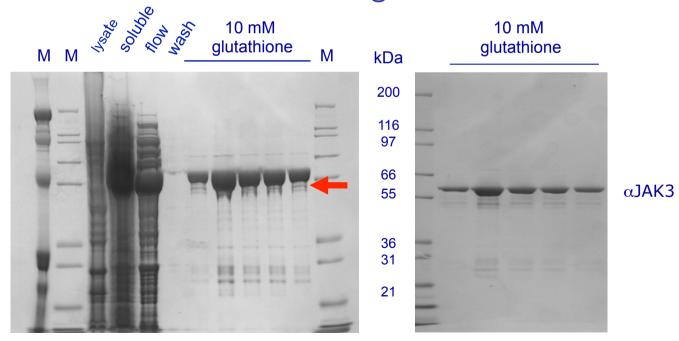
#### ▶ LC-MS on C4 column with MeCN/water (0.05% TFA)



## Tyrosine Kinase JAK3

- Kinase domain expressed as ca. 60 kDa GST-fusion protein in E. coli:

   GST
   LVPGS
- One-step purification on immobilized glutathione
- Analytical SEC and DLS: homogenous, ca. 800 kDa
- SDS-PAGE and immunoblotting:



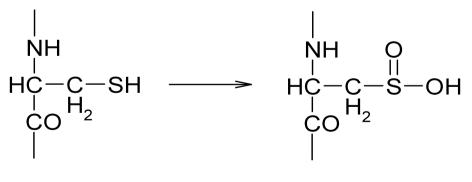


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- Secondary + tertiary structure
  - CD, FTIR
  - Thermal unfolding (DSC, CD, ThermoFluor<sup>®</sup> Assay)
  - Specific activity of enzymes, [ITC, CFCA Assay on SPR]



## Cysteine Proteinase Z

- 55 kDa proenzyme refolded from E. coli inclusion bodies, using factorial design
- Acid-induced autocatalytic cleavage and isolation by cation exchange chromatography
- Homogenous by SDS page
- Oxidation of active-site cysteine detected by Xray crystallography



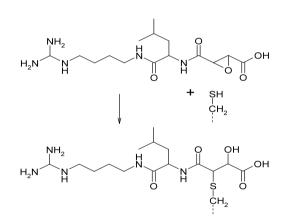
 $pK_{s} = 8-9$ 

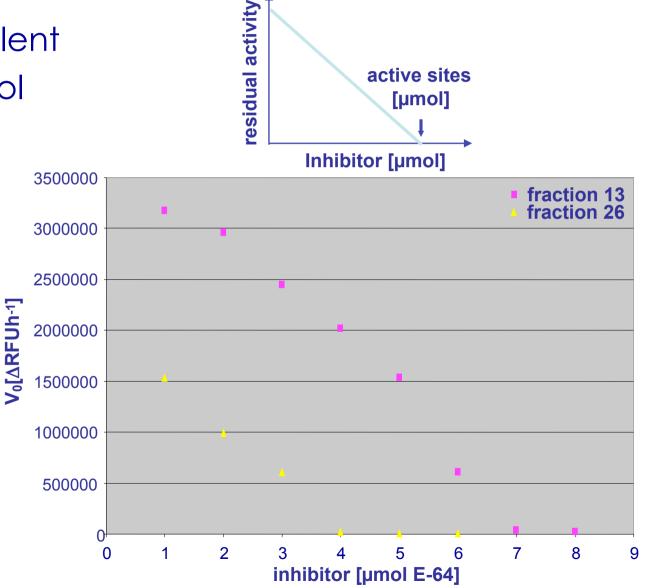
$$pK_{s} = 3-4$$



## Active Site Titration

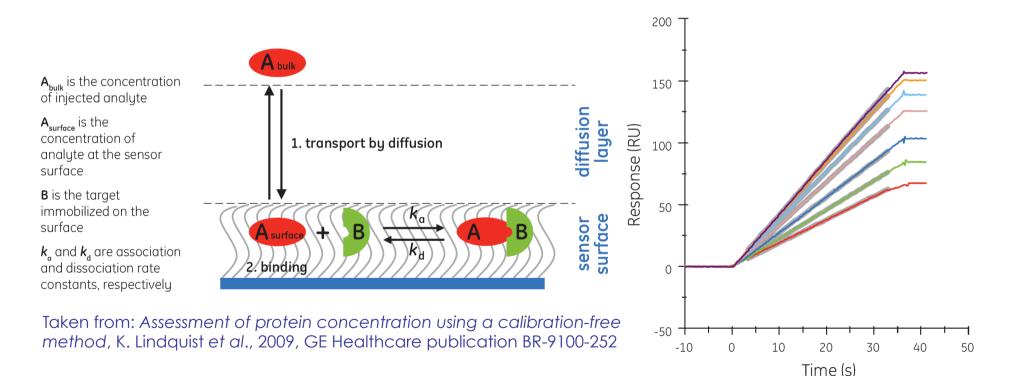
• E-64 is a covalent inhibitor for thiol proteases





## SPR/Biacore-based methods

### Example for an SPR-based assay : Calibration Free Concentration Assay CFCA



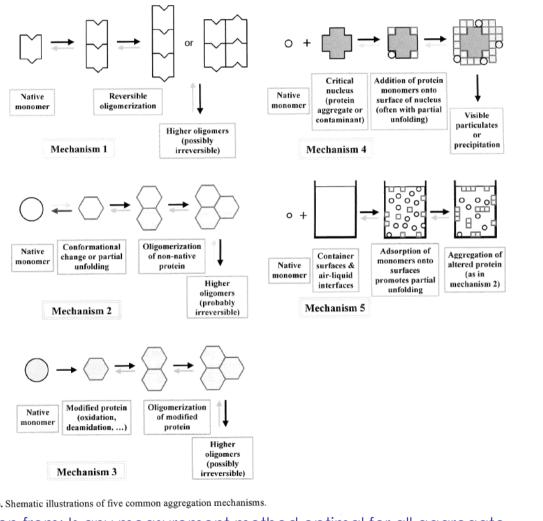


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- Secondary + tertiary structure
  - CD, FTIR
  - Thermal unfolding (DSC, CD, Therms)
  - Specific activity of enzymes, [ITC, CFC Assay on SPR]
- Quarternary Structure: Oligomerization state + aggregation
  - DLS, SLS, SEC-MALS, AUC [UV, fluorescence, dye staining] etc.



## What is aggregation?



### But the most neglected aspect of aggregation is kinetics!

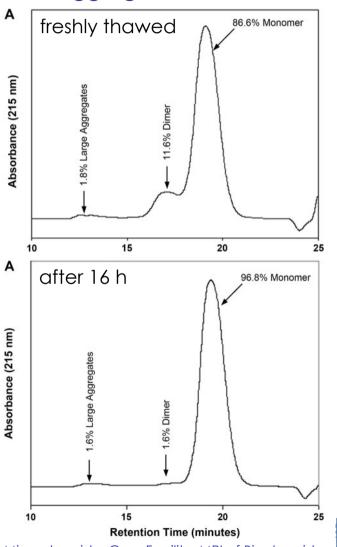
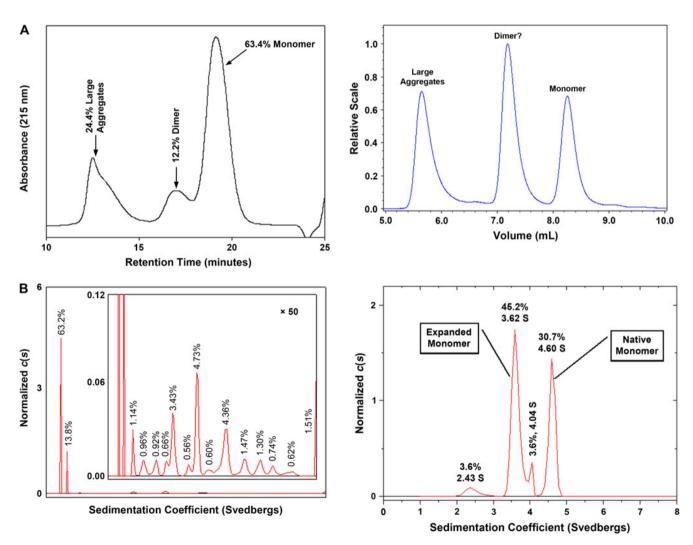


Fig. (1). Shematic illustrations of five common aggregation mechanisms.

Taken from: Is any measurement method optimal for all aggregate sizes and types? John Philo, AAPS Journal 2006, E564ff.

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## Usefullness of SEC as standard



Dissociation or loss of aggregates can be caused by:	SEC	SV	FFFF
dilution	+++	+	+++
change of solvent conditions	+++	_	++
adsorption to surfaces	+++	+	++
physical filtration (eg, column frit)	+++	_	_
physical disruption (eg, shear forces)	++	_	_
Creation of new aggregates can be caused by:			
change of solvent conditions	+++	_	++
surface or shear-induced denaturation	++	_	+
concentration on surface	_	_	+

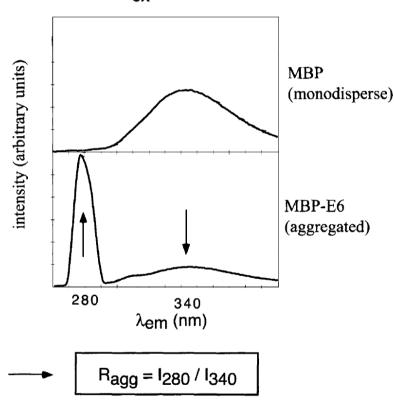
\*The number of pluses indicates the relative size of the problem for that method.

SEC indicates size exclusion chromatography; SV, sedimentation velocity; FFFF, flow field-flow fractionation.



### Usefullness of UV and Fluorescence

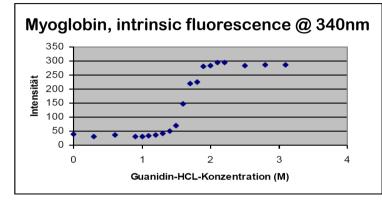
#### Aggregation index



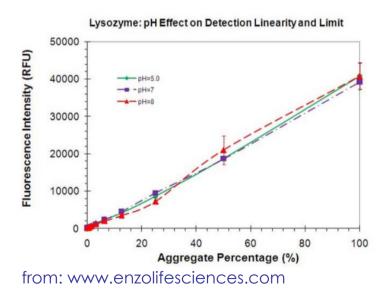
λ<sub>ex</sub> = 280 nm

Nominé et al., Protein Engineering 2006, 297ff.

#### But what about quantum efficiency?



#### Keller, Uni Graz



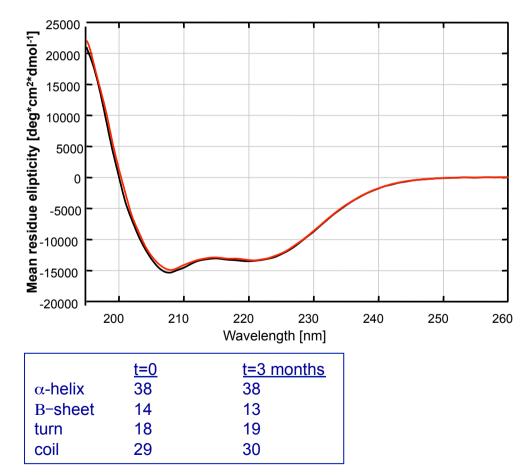


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  - [peptide mapping by MS of tryptic digests]
- Secondary + tertiary structure
  - CD, FTIR
  - Thermal unfolding (DSC, CD, ThermoFluor® Assay)
  - Specific activity of enzymes, [ITC, CFCA Assay on SPR]
- Quarternary Structure: Oligometization state + aggregation
  - DLS, SLS, SEC-MALS, AUC [UV, Dece, dye staining] etc.
- Protein stability
  - Aggregation (and activity) as surrogate markers of stability



## Matrix Metalloproteinase 12

Protein stability investigated using CD spectroscopy:





- Identity i.e. primary structure: amino acid sequence and desired posttranslational modifications
  - Edman sequencing combined with protein total mass (ESI-TOF, ESI-FTMS, [MALDI(IR)-TOF], deglycosylation of secreted proteins from insect cells and S.cerevisiae)
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- Quarternary Structure: Oligomerization state + aggregation
  - DLS, SLS, SEC-MALS, AUC [UV, fluorescence, dye staining] etc.
- Protein stability
  - Aggregation (and activity) ?? ?% Ate markers of stability
- Purity
  - Cave: lack of linearity in staining

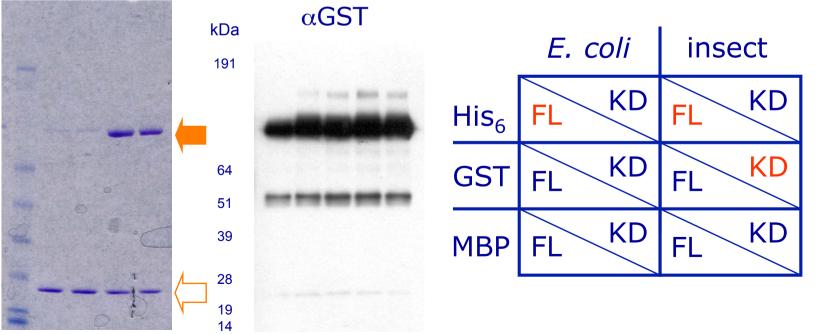


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  - [peptide mapping by MS of tryptic digests]
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- Protein stability
  - Aggregation (and activity) as surrogate markers of stability
- Purity
  - Cave: lack of linearity in staining



## Tyrosine Kinase BTK

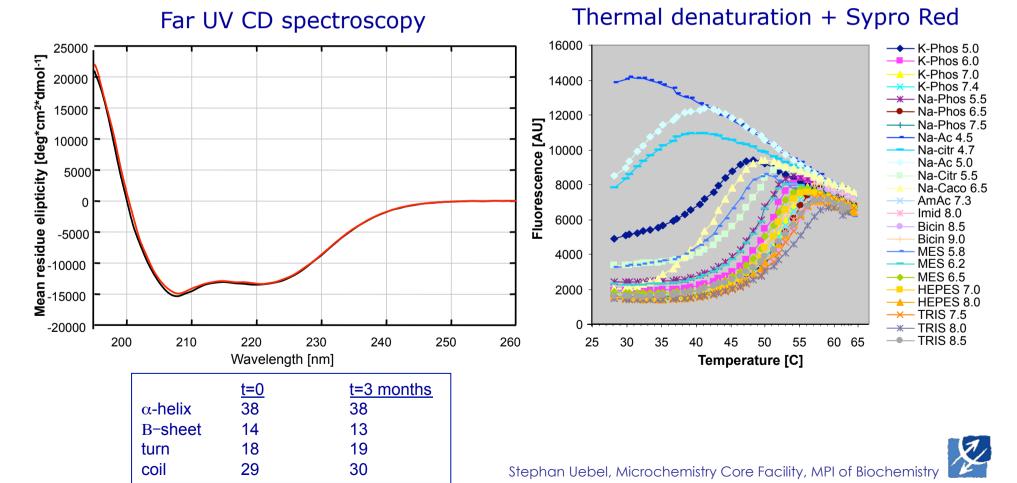
- Kinase domain expressed as ca. 78 kDa GST- fusion protein in insect cells
- One-step purification on immobilized glutathione
- Contaminant at ca. 25 kDa recognised by αGST antibody, but not identified by MALDI-TOF MS



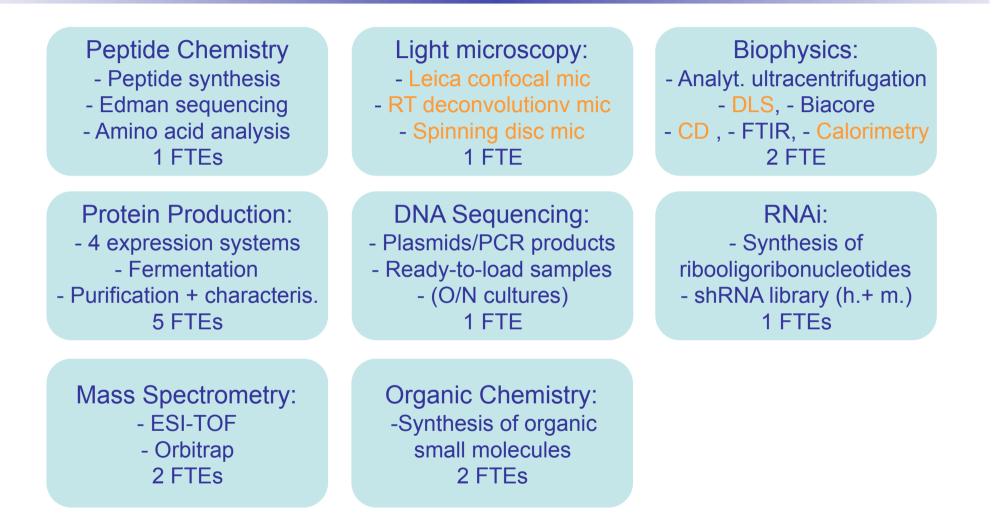


# Matrix Metalloproteinase 12

Protein stability is investigated using CD spectroscopy and fluorescent dye binding



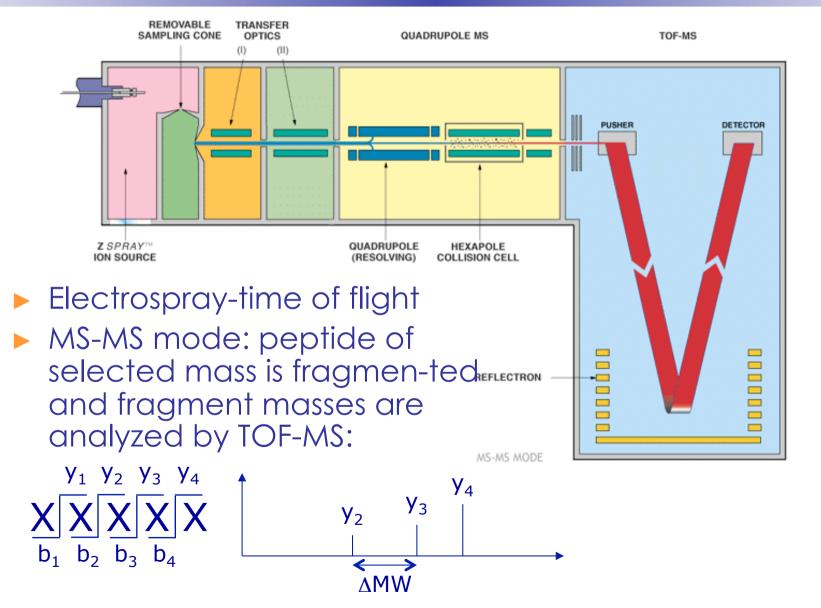
## Overview



Total: 16 full time employees (five scientists); open access instruments in orange

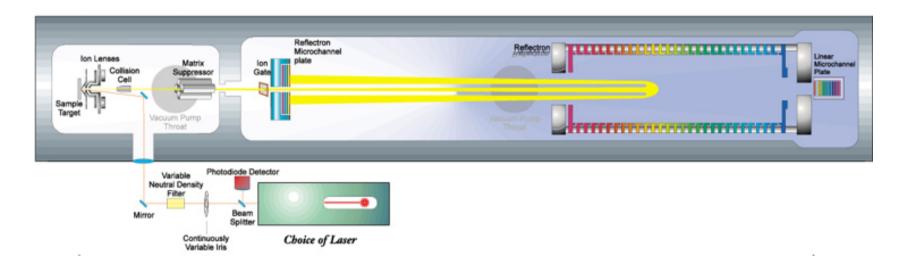


### ESI-TOF Tandem MS MS





## MALDI- TOF MS of Tryptic Digests



- Matrix assisted laser desorption ionization time of flight mass spectrometry
- Peptides are embedded in a UV-absorbing matrix
- The list of peptide masses is run against a theoretical list of peptide masses after tryptic digest of proteins in the database of choice (e.g. SWISSPROT)



## Analytische UZ

- Sedimentation velocity is an analytical ultracentrifugation method that measures the rate at which the sedimentation boundary moves in response to centrifugal force. It measures the sedimentation coefficient (S) which depends on the molecular weight (larger proteins sediment faster) and also on molecular shape.
- Sedimentation equilibrium measurements provide thermo-dynamically rigorous, shape independent, molecular weights:
  - As the centrifugal force produces a gradient in protein concentration across the centrifuge cell, diffusion acts to oppose this concentration gradient. Eventually, an exact balance is reached between sedimentation and diffusion, and the concentration distribution reaches an equilibrium. The concentration distribution at equilibrium depends only on molecular mass, and is entirely independent of the shape of the molecule.

